Analogues of Ornithine as Inhibitors of Ornithine Decarboxylase. New Deductions Concerning the Topography of the Enzyme's Active Site

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Fourteen structural analogues of ornithine were synthesized and evaluated as inhibitors of preparations of the enzyme L-ornithine carboxylyase (ODC) (E.C. 4.1.1.17) obtained from rat liver, rat hepatoma cells in culture, or bull prostate. The synthesis of these compounds was achieved either via a Bucherer type reaction or via alkylation of carbanions derived from ethyl acetamidocyanoacetate, methyl isocyanoacetate, benzyl α -isocyanopropionate, methylbenzaldimine alanate, and the azlactone derivative of ornithuric acid. (+)- α -Methylornithine, which was assigned the L configuration on the basis of rotational criteria, was found to be the most potent reversible inhibitor of ODC among the synthesized compounds. From the degree of inhibition of ODC activity in the presence of the various ornithine analogues, it has been possible to delineate some of the structural features of the substrate L-ornithine which are required for binding to the mammalian ODC active site.

Numerous investigations clearly indicate that the polyamines, spermidine and spermine, and their diamine precursor, putrescine, have essential functions in cellular mechanisms, particularly those involved in cell division.¹ A possible way of elucidating the biological roles of these bioamines is to selectively impair their biosynthesis.² In eukaryotic organisms the conversion of ornithine to putrescine catalyzed by ornithine decarboxylase (ODC) (Lornithine carboxylyase E.C. 4.1.1.17) is believed to be the obligatory and rate-limiting step for their biosynthesis,3 a concept that has led to the search for specific inhibitors of this enzyme. Thus far, the substrate analogues 2hydrazino-5-aminopentanoic acid, DL- α -methylornithine, δ DL-2-hydrazino-2-methyl-5-aminopentanoic acid, ⁶ N-(5'phosphopyridoxyl)ornithine, and DL-(E)-2,5-diamino-3-pentenoic acid⁸ have been found to be potent reversible inhibitors of ODC.

We are engaged in a program for the design of specific inhibitors of ODC and report here on some steric, electronic, and conformational features of ornithine analogues essential for their function as inhibitors of ODC. Fourteen structural analogues of ornithine were synthesized and used to probe the specificity of L-ornithine binding sites on ODC. These compounds were selected on the basis of (a) conformational restrictions resulting from ring formation, (b) different substitution patterns on the terminal amine function and/or on the α - and β -carbon atom, (c) modifications of the carboxyl group, and (d) different configurations of the α -carbon atom. The effect of these molecular characteristics has been studied with the crude extract of ODC obtained from rat liver, bull prostate, and rat hepatoma (HTC) cells in culture.

Results and Discussion

Chemistry. The acyclic analogues of ornithine synthesized for this study are listed in Table I. a-Hydroxymethylornithine 7 was prepared in good overall yield from ornithuric acid 15 by the general method described by Kaminski et al. 11 for the synthesis of α -hydroxymethyl- α -amino acids from the corresponding α -amino acids (Scheme I). β -Hydroxyornithine 9^{12} was synthesized by alkylation of the anion derived from ethyl isocyanoacetate¹³ with β-phthalimidopropanal, ¹⁴ followed by hydrolytic removal of the protective groups, according to the procedure reported by Schöllkopf and Hoppe¹⁵ for the preparation of β -hydroxy- α -amino acids. The synthesis of the α -methylornithine analogues 1–6, 11, and 12 relied on the utilization of either the α -isocyanopropionate synthon 25¹⁶ or the Schiff base methyl alanate synthon 29.17 Alkylation of the anion derived from benzyl α -isocyanopropionate (25) with (N-methyl-N-tosyl)-3-iodo-1propanamine (26), 18 N,N-dimethyl-3-chloro-1-propanamine

(27), ¹⁹ and [N-tosyl-N-(3-acetamidopropyl)]-3-chloro-1propanamine (28), in the presence of a catalytic amount of sodium iodide in the two latter cases, gave in good yield the corresponding C-alkylated products 16a-c (Scheme II). Hydrolytic removal of the ester and amide protective groups from 16a-c, followed by sodium ammonia reductive cleavage of the N-tosyl bond, afforded the corresponding α -methylornithine derivatives 1-3. The halide 28 was prepared by reacting 3-(N-2'-cyanoethyl)amino-1propanol²⁰ with 2 equiv of tosyl chloride in pyridine, followed by catalytic reduction of the nitrile in acetic anhydride. The Schiff base methyl alanate synthon 29, in addition to being more conveniently available than the α -isocyanopropionate synthon 25, allows a discrimation in the deprotection of the carboxyl and amine functions. This property has been used to advantage for the preparation of the key piperidone intermediate 18 (Scheme III) in the synthesis of DL- α -methylornithine 6 and its enantiomers 4 and 5. Alkylation of the anion of the Schiff base methyl alanate with N-(3-iodopropyl)phthalimide proceeded smoothly to give an excellent yield of 17a. Mild acidic hydrolysis of 17a in 1 M HCl for 1 h removed the benzylidene group to give 19 which upon treatment with 1 equiv of hydrazine in the presence of 1 equiv of sodium methylate in methanol afforded quantitatively the piperidone 18 which has been easily resolved via its (+)- and (-)-binaphthylphosphoric salt.²¹ Hydrolysis of the enantiomeric piperidone binaphthylphosphoric salt yielded

Table I. Analogues of Ornithine Synthesized as Inhibitors of ODC

the corresponding α -methylornithine enantiomers. The L configuration was assigned to the (+) enantiomer 4 of α -methylornithine on the basis of the following rotational criteria. (a) The rotatory dispersion curve of 4 is similar to that of L-(+)-ornithine. The rotation of both amino acids becomes strongly positive in the 350-nm region (Table II). (b) The Lutz-Jirgensons rule²² that L- α -amino acids show a positive shift in rotation upon increasing the acid concentration of the solution, which has been demonstrated by Achiwa et al.23 to apply also to L-\alphamethyl- α -amino acids, is obeyed by 4. The specific rotation difference $[\alpha]$ (6 M HCl) – $[\alpha]$ (0.1 M HCl) of 4 is positive at all wavelengths. The same specific rotation difference measured for L-(+)-ornithine follows a similar pattern (Table II). (c) L- α -Amino acids and L- α -methyl- α -amino acids give hydantoin derivatives with large negative rotation. According to Achiwa et al.,23 this is the most reliable rule for assigning the absolute configuration of α -methyl- α -amino acids. The hydantoin derivative 30 of (+)- α -methylornithine 4 was synthesized and a negative rotation was measured ($[\alpha]^{20}_D$ -14.5°). The higher and the lower homologues of α -methylornithine, 11 and 12, were prepared in a similar manner using the higher and the lower homologues of N-(3-iodopropyl)phthalimide as alkylating agents. The synthesis of the ornithine analogues 8 and 10 was approached through the alkylation of the ethyl acetamidocyanoacetate synthon²⁴ with respectively 28 and N-(3-iodopropyl)phthalimide. Hydrolytic removal of the protective groups and reductive cleavage of the

Table II. Influence of Wavelength on Optical Rotation and on Shift in Rotation Induced by Acid Concentration on D- and L-Ornithine and on (+)- and (-)- α -Methylornithine

λ, n m	$[\alpha]$, deg $(c 2, H_2O)$				
	4	L- Ornithine	5	D- Ornithine	
58 9	+8.8	+11.5	-10.2	-11.7	
578	+9.4	+12.1	-10.5	-12.4	
546	+10.5	+13.9	-11.9	-14.1	
436	+18.2	+24.7	-20.5	-25.2	
365	+29.0	+42.1	-32.8	-42.8	
	[\alpha] (6 M	$HCl) - [\alpha](0$.1 M HCl), deg (c 2)	
		T -		D-	

λ, nm	4	L- Ornithine	5	D- Ornithine	
589	+2.0	+ 5.4	-2.0	-3.5	
578	+1.8	+5.5	-1.9	-3.4	
546	+1.6	+6.5	-2.3	-3.8	
436	+3.8	+11.5	-3.6	-7	
365	+6.5	+19.4	-5.8	-12.2	

Scheme IV

N-tosyl bond of the C-alkylated product 20 afforded the aminopropylornithine derivative 8, whereas treatment of the C-alkylated product 21 with sodium azide in dimethylformamide gave the tetrazolyl derivative 22 which upon boiling in concentrated HCl and chromatography on ion-exchange resin yielded the tetrazolylornithine analogue 10 (Scheme IV). The cyclic analogues 13 and 14 were

Table III. Kinetic Constants $K_{\rm I}$ (mM) for ODC Inhibition by Ornithine Analogues

Compd	Rat liver	HTC cell	Bull prostate
L-Ornithine	0.087ª	0.09ª	0.072ª
$egin{array}{c} 1 \ 2 \end{array}$	0.5^{b}		0.74
	No inhibn at 1 mM		
3		No inhibn	4.4
	0.019	0.02	0.02
5	1.3	1.05	1.0
6	0.04	0.04	
4 5 6 7	No inhibn at 1 mM		
8			5.5
9		No inhibn at 1 mM	
10	0.12	0.14^{b}	0.07
11	No inhibn at 4 mM		
12	No inhibn at 4 mM		
13	0.07	0.07	
14	No inhibn at 0.25 mM		

 a Corresponds to the $K_{\rm M}$. b These values were calculated from the ID $_{\rm so}$ measured at an ornithine concentration equal to the $K_{\rm M}$. Under these conditions, for a competitive inhibitor ID $_{\rm so}$ = 2 \times $K_{\rm I}$ (ref 36).

obtained via a route essentially identical with the one described by Christensen and Cullen²⁵ except that 4-acetamidocyclohexanone instead of 4-p-toluenesulfon-amidocyclohexanone was used as starting material. The stereochemistry of the 4-amino group in compounds 13 and

14 was established by treating the isomers separately at 300 °C for 1 min in a sealed tube. The *trans*-1,4-diaminocyclohexane-1-carboxylic acid 13 gave the bicyclic lactam 4-amino-2-azabicyclo[2.2.2]octan-3-one, whereas the cis isomer 14 remained unchanged.

Enzyme Inhibitory Activity. The structural analogues of ornithine synthesized as part of this study were tested as competitive inhibitors of ODC. Rat liver, hepatoma cells in culture, and bull prostate were used as the source of enzyme. All three enzyme preparations had the same $K_{\rm M} \simeq 0.08$ mM for L-ornithine. An apparent inhibition at high concentration of the substrate was observed with bull prostate ODC. Such an inhibition has already been reported for rat prostate ODC⁴ and for mouse liver ODC. ²⁶

The degree of inhibition of ODC activity by the structural analogues of ornithine expressed by their $K_{\rm I}$ is shown in Table III. (+)- α -Methylornithine 4 was the most effective inhibitor with a $K_{\rm I}=0.02$ mM for the three different ODC preparations. Its inhibitory activity was twice that of the racemic mixture 6. Its antipode 5 displayed a very low inhibitory activity ($^1/_{70}$ th of that of 4). This finding is an agreement with the results of Ono et al. who showed that D-ornithine is a very weak inhibitor of rat prostate ODC (25% inhibition at a concentration of 10 mM) and of Inoue et al. Who demonstrated that rat prostate and liver ODC have much less affinity for D-2-hydrazino-5-aminopentanoic acid than for L-2-hydrazino-5-aminopentanoic acid and, therefore, confirms the

preferred L configuration of the ligand for binding to the enzyme's active site.

A primary terminal amino group in the ligand appears to be important for enzyme inhibition, since the δ -Nmonomethyl- and the δ -N,N-dimethyl- α -methylornithine analogues 1 and 2 showed little or no inhibitory activity on rat liver ODC. Furthermore, the δ -N-aminopropyl derivatives of ornithine and of α -methylornithine 8 and 3, which incorporate in the same molecule the spermidine and the ornithine or α -methylornithine skeletons, proved also to have a very low affinity for bull prostate ODC (K_1 = 5.5 and 4.4 mM), despite the fact that spermidine was shown to be a weak competitive inhibitor of bull prostate ODC with a $K_1 = 3$ mM, comparable to the value ($K_1 =$ 2.7 mM) already reported for rat prostate ODC.28 This result suggests that spermidine and ornithine compete for the same binding site of prostate ODC and also that the enzyme cannot accommodate ligands with bulky groups on the terminal amino group.

Of the two cyclic ornithine analogues, only the trans-1,4-diaminocyclohexane-1-carboxylic acid (13) is a potent inhibitor of ODC. Its $K_I = 0.07$ mM is 3.5 times higher than that of (+)- α -methylornithine 4. The cis isomer 14 is devoid of activity. These findings which indicate that the distance between the two nitrogen atoms of the ornithine analogues is of major importance for inhibition are consistent with the fact that neither the lower and the higher homologues of ornithine nor their corresponding α -methyl analogues 11 and 12 are substrates or inhibitors of ODC. This suggests that ornithine adopts a fully stretched conformation in the enzyme's active site with the two nitrogen atoms as far away as possible, which according to the Dreiding model corresponds to a distance of about 6 Å. Most probably hydrophobic interactions with residues of the active site favor this stretched conformation. This assumption is corroborated by the high affinity for ODC displayed by DL-(E)-2,5-diamino-3pentenoic acid. The presence of the trans double bond results in a restricted number of possible conformers, most of them displaying the favorable distance of about 6 Å between the two nitrogen atoms, and also in an enhanced polarization when compared to the saturated analogue, thus leading to a higher interaction energy with the residues of the enzyme's active site via hydrophobic bonding. Relyea and Rando⁸ reported for this inhibitor a $K_{\rm I}$ of 4.4 \times 10⁻⁶ M for rat liver ODC, that is to say an affinity for the enzyme 30 times higher than that of DL-ornithine. Introduction of a hydrophilic substituent such as a hydroxyl function on the β -carbon atom, analogue 9, or on the α -methyl group, analogue 7, completely abolished the affinity for the enzyme's active site. The major contribution to this loss of affinity cannot be steric hindrance to binding, since the β -methyl-²⁹ and the α -ethylornithine³⁰ analogues have been reported to retain some inhibitory activity, but is rather due to a disruption by the hydrophilic function of the hydrophobic interactions between the residues of the enzyme's active site and the inhibitors.

Replacement of the carboxyl group of the substrate by a tetrazolyl group did not decrease the affinity for ODC, as shown by the $K_{\rm I}=0.07$ mM measured for the (±)-tetrazolyl analogue 10 vs. a $K_{\rm M}=0.07$ mM for L-ornithine for bull prostate ODC. If we assume that only one enantiomer of the tetrazolyl analogue 10 is accepted by the enzyme's active site, as suggested by the inhibitory activities of (+)- and (-)- α -methylornithine, the abovementioned replacement results in fact in a slightly increased affinity for the binding sites. The p $K_{\rm a}$ values of the various functions of the tetrazolyl analogue 10 were

Table IV. Comparison of the Stability Constants K_d for the Formation of the Schiff Base with PLP and the $K_{\rm I}$ of Some ODC Inhibitors

	L- Orni- thine	1	3	4	Sper- mi- dine	Pu- tres- cine
K _d , mM	1.25	5.36	0.9	2.35	0.7	0.8
$K_{ m I}, \\ m mM$	0.07	0.74	4.4	0.02	3	

found to be in the same range as those of the corresponding functions of ornithine (2.95, 7.80, and 10.25, respectively, for the tetrazolyl, the α -amino, and the terminal amino group of 10 vs. 2.8, 8.75, and 10.4 for the corresponding functions of L-ornithine). One interpretation of these data is that the carboxyl group is an attachment point via electrostatic binding for the ligand and that the area on the enzyme which accommodates the carboxyl group is not sensitive to changes in the steric bulk of the ligand. An alternative explanation suggested by the poor affinity of putrescine for the enzyme's active site is that the binding of the ligand is highly dependent on the basicity of the α -amino group and that the carboxyl or the tetrazolyl group would not participate directly in the binding but would act by decreasing the p K_a of the α -amino group.

It was of importance to ascertain that the affinity of the ligand for the enzyme was not dependent on its affinity for the cofactor. For this purpose, the dissociation or stability constants K_d for the formation of the Schiff base with pyridoxal phosphate (PLP) of some ornithine analogues and of spermidine and putrescine were determined and compared to the $K_{\rm I}$ of the same analogues for ODC. The results summarized in Table IV undoubtedly show that there is no correlation between the K_d and the K_I and therefore rule out any important role for the ligand affinity for the PLP in its binding to the enzyme.

Conclusion

The delineation of the structural features of L-ornithine which are required for binding to mammalian ODC was approached by measuring the degree of inhibition of ODC activity in the presence of various structural analogues of ornithine. The requirement for an L configuration of the ligand for binding has been confirmed. It has also been demonstrated that (1) a distance between the two nitrogen atoms of the ligand of about 6 Å is crucial for binding; (2) the area of the enzyme which accommodates the terminal amino group of the substrate is very sensitive to steric hindrance, whereas that which accommodates the carboxyl group is not; (3) hydrophobic interactions with the side chain of the ligand are an important feature for the binding, since hydrophilic functions attached on the α - and the β -carbon atoms abolish all affinity for the enzyme; and (4) the affinity of the ligand for the enzyme is not directly dependent on its affinity for the PLP cofactor.

Experimental Section

Enzyme Purification. Bull Prostate ODC. The enzyme was purified about fourfold according to the procedure of Jänne and Williams-Ashman³¹ with some modifications. A fresh bull prostate was rapidly dissected free of surrounding connective tissue, weighed, minced, and homogenized in 2 vol of homogenization medium (Tris-HCl buffer, pH 7.2, 10 mM; sucrose, 25 mM; tetrasodium EDTA, 0.1 mM; dithiothreitol, 5 mM; PLP, 0.1 mM) with a Waring blendor and then with a glass homogenizer. All operations were carried out at 4 °C. The homogenate was centrifuged at 20000g for 30 min at 2 °C. The resulting supernatant fraction was passed through a cheesecloth in order to remove floating lipids. This fraction was centrifuged at 105 000g for 1 h. The supernatant fluid was fractionated with solid

(NH₄)₂SO₄. The fraction precipitating between 20 and 50% of saturation was dissolved in 6 mL of conservation medium (homogenization medium minus sucrose) and dialyzed against 100 vol of the same buffer for 16 h, the dialyzing buffer being changed after the first 6 h. The dialyzed (NH₄)₂SO₄ fraction was divided into small aliquots and stored at -20 °C.

Rat Liver ODC. This was prepared from the livers of rats which had been injected with thioacetamide 18 h before sacrifice and was purified about tenfold by acid treatment at pH 4.6 as described by Ono et al.9

HTC Cell ODC. The preparation was obtained as described by McCann et al.¹⁰

Assay of Ornithine Decarboxylase Activity. The reactions were followed by measurement of release of 14CO2 from DL-[1-14C]ornithine by the method of Jänne and Williams-Ashman.31 The standard incubation mixture contained in a final volume of 0.95 mL: 30 µmol of Tris-HCl, pH 7.2 (bull prostate ODC), or sodium phosphate buffer, pH 7.1 (HTC and rat liver ODC), 0.1 μ mol of pyridoxal 5-phosphate, 5 μ mol of dithiothreitol, 0.043 μ mol of DL-[1-14C] ornithine (58 Ci/mol), and 0.8 µmol of L-ornithine. The reaction was initiated by addition of 50 μ L of enzyme solution. Release of ¹⁴CO₂ was measured after 1 h of incubation at 37 °C. Under these conditions, the activity of ODC preparations from bull prostate, rat liver, and HTC cells was respectively 4.8, 7.2, and 7.5 mmol of CO₂/h per mg of protein.

Data Processing. Kinetic constants were calculated from Lineweaver-Burk plots $(K_{\rm M})^{32}$ or from Dixon plots $(K_{\rm I})^{33}$ Dissociation constants $(K_{\rm d})$ of PLP adduct with original plots $(K_{\rm I})^{33}$ analogues were determined at 25 °C in a phosphate buffer (30 mM, pH 7.1) by the method of Lucas et al.³⁴ All inhibition constants were obtained from a least-squares fit of the data points with a Hewlett-Packard 9820 A calculator (accuracy better than

Chemistry. Melting points were obtained on a Mettler FP5 or a Büchi SMP-20. Microanalyses were conducted on a Perkin-Elmer 240 CHN analyzer. Unless otherwise stated, the IR, NMR, and UV data were consistent with the assigned structure. IR data were recorded on a Beckman IR-577 or Beckman IR 257 spectrophotometer, NMR data on a Varian Associates Model T-60 spectrophotometer (Me₄Si), and UV data on a Perkin-Elmer 402 spectrophotometer.

5-Benzoylamino-5-(3-benzoylaminopropyl)-1,3-dioxan-4-one (23). A mixture of 2-phenyl-4-(3-benzoylaminopropyl)-4,5-dihydro-1,3-oxazol-5-one (24) (25 g, 0.0775 mol) (prepared by treatment of ornithuric acid with acetic anhydride at 90 °C for 0.5 h), pyridine (17 mL), and formaldehyde (57 mL of a 37% aqueous solution) was stirred at ambient temperature for 20 h. The reaction mixture was then quenched with water (50 mL). The white solid which precipitated was filtered, washed with water and then with petroleum ether, and dried over P₂O₅ to give a white powder, 24.18 g (81%), mp 190-193 °C, which could be used without further purification. The analytical sample was recrystallized from acetone-petroleum ether, mp 196-197 °C. Anal. $(C_{21}H_{22}N_2O_5)$ C, H, N.

α-Hydroxymethylornithine Monohydrochloride (7). To a suspension of 23 (24 g, 0.063 mol) in anhydrous MeOH (100 mL) was added a catalytic amount of NaOMe. After stirring for 0.25 h, the reaction mixture which had become homogeneous was neutralized with concentrated HCl. The solvent was evaporated under reduced pressure and the residue extracted with CH₂Cl₂. The organic phase was washed with brine and then concentrated in vacuo to leave an oily residue (24.12 g) which was treated in 6 M HCl (150 mL) at reflux for 48 h. The benzoic acid which precipitated on cooling was separated and the filtrate was extracted with ether (4 × 100 mL). The aqueous phase was concentrated under reduced pressure. The residue was dissolved in absolute ethanol (180 mL) and the pH of the solution was adjusted to 4 by adding a solution of triethylamine in ethanol (1 M) (56 mL). The solid which formed was collected by filtration, washed with ethanol, and recrystallized (H₂O-EtOH) to yield 10.4 g (87%) of 7, mp 246 °C. Anal. (C₆H₁₅N₂O₃Cl) C, H, N.

Methyl 2-Methyl-2-benzylideneamino-5-phthalimidopentanoate (17a). To a solution of lithium diisopropylamine (0.01 mol) in anhydrous THF (15 mL) [prepared in situ from a solution of diisopropylamine (1 M) in THF (0.01 mol) and a solution of butyllithium (2 M) in hexane (0.01 mol)] cooled to

 $-78~^{\circ}\mathrm{C}$ was added slowly under nitrogen a solution of methylbenzaldimine alanate (29) (1.91 g, 0.01 mol) in anhydrous THF. The reaction mixture turned red immediately. The temperature was allowed to rise to $-20~^{\circ}\mathrm{C}$ and then a solution of N-(3-iodopropyl)phthalimide (3.94 g, 0.0125 mol) in anhydrous THF (30 mL) was added. Stirring was continued for 4 h while the temperature was allowed to rise to $20~^{\circ}\mathrm{C}$. The reaction mixture was quenched with water. The organic phase was separated and the aqueous phase was extracted with ether (2 \times 100 mL). The combined organic phases were washed with brine, dried over MgSO₄, and concentrated in vacuo to yield an oily residue of crude 17a (3.6 g).

Methyl 2-Methyl-2-benzylideneamino-6-phthalimidohexanoate (17b). Crude 17b was obtained in a manner similar to that described for 17a. N-(4-Iodobutyl)phthalimide was used in place of N-(3-iodopropyl)phthalimide.

Methyl 2-Methyl-2-benzylideneamino-4-phthalimidobutanoate (17c). Crude 17c was obtained in a manner similar to that described for 17a except that HMPA was added as a cosolvent. N-(2-Iodoethyl)phthalimide was used in place of N-(3-iodopropyl)phthalimide.

DL- α -Methylornithine Hydrochloride Monohydrate (6). Crude 17a (3.6 g) was suspended in 6 M HCl (50 mL) and heated at reflux for 4 days. The phthalic acid which precipitated on cooling was filtered off and the filtrate was extracted with CHCl₃. The aqueous phase was concentrated under reduced pressure. The remaining phthalic acid which precipitated was eliminated by filtration. The residue was then dissolved in absolute ethanol and the pH of the solution adjusted to 4 with a solution of triethylamine in ethanol (1 M). The solid which formed was collected by filtration, washed with CHCl₃, and recrystallized (H₂O–EtOH) to yield 1.16 g of analytically pure α -methylornithine hydrochloride monohydrate, mp 255 °C. Anal. ($C_6H_{17}O_3N_2Cl$) C, H, N.

DL- α -Methyllysine Hydrochloride Monohydrate (11). Treatment of crude 17b in a manner similar to that described for 17a afforded 11, mp 266 °C. Anal. ($C_7H_{19}N_2O_3Cl$) C, H, N.

DL-2-Methyl-2,4-diaminobutyric Acid Hydrochloride (12). Treatment of crude 17c in a manner similar to that described for 17a afforded 12, mp 224 °C. Anal. (C₅H₁₃N₂O₂Cl) C, H, N.

 δ -(N-Phthaloyl)- α -methylornithine Methyl Ester Hydrochloride (19). 17a (700 g) was stirred vigorously for 3 h in aqueous 1 M HCl (4 L). The oily phase which separated was eliminated by decantation and the aqueous phase was washed with petroleum ether (3 × 1.5 L) and then concentrated in vacuo. The solid which precipitated was collected by filtration, washed with petroleum ether and CCl₄, and then dried in vacuo to yield 479 g of 19. The analytical sample was obtained by recrystallization from H₂O: mp 243 °C. Anal. (C₁₅H₁₉N₂O₄Cl) C, H, N.

3-Amino-3-methyl-2-piperidone (18). To a suspension of 19 (469 g, 1.44 mol) in anhydrous MeOH (1 L) was added under N_2 a solution of NaOMe (33 g of Na, 1.44 g-atoms) in MeOH (1 L), followed by a solution of hydrazine hydrate (72 g, 1.44 mol) in MeOH (0.5 L). The reaction mixture was heated at reflux for 7 h. The solid which formed was separated by filtration and the filtrate was concentrated in vacuo. The residue was treated with hot CHCl₃ and the insoluble part (22 g) was discarded. Evaporation of the solvent afforded 178 g of 18 (97.5%), mp 142 °C. Anal. (C₆H₁₂N₂O) C, H, N.

(+)- α -Methylornithine Hydrochloride (4). To a solution of 18 (3.07 g, 0.0236 mol) in ethanol (50 mL) maintained at 50 °C was added a boiling solution of (-)-binaphthylphosphoric acid (8.2 g, 0.0236 mol) in ethanol (250 mL). The reaction mixture was maintained at 60 °C until crystallization started. Thereafter, it was allowed to cool slowly to room temperature and then maintained at 4 °C for 1.5 h. The crystals were collected by filtration and washed with ethanol (50 mL) and ether (2 × 100 mL) to give 5.47 g of the (-)-binaphthylphosphoric salt (95%) [mp 336 °C dec; $[\alpha]^{20}_D$ -432° (c 0.9, MeOH). Anal. (C₂₆H₂₅N₂O₅P) C, H, N], which was then heated at reflux temperature in 6 M HCl (40 mL) for 2 h. The solid which separated was filtered and washed with water (3 \times 10 mL), and the filtrate was concentrated in vacuo. The crystalline residue was dissolved in water (~ 10 mL) and the pH of the solution was adjusted to 3.5 by adding triethylamine. Concentration under reduced pressure yielded a solid which was treated with hot $CHCl_3$ (3 × 10 mL). The insoluble residue was then crystallized (H₂O–EtOH) to yield 1.7 g of 4 (80%): mp 218 °C; $[\alpha]^{20}_D$ +8.4° (c 0.5, H₂O). Anal. (C₆H₁₅N₂O₂Cl) C, H, N.

(-)- α -Methylornithine hydrochloride (5) was obtained in a manner similar to that described for 4. (+)-Binaphthylphosphoric acid was used in the place of the (-) enantiomer: mp 214 °C; $[\alpha]^{20}_D$ -10° (c 1, H₂O). Anal. (C₆H₁₅N₂O₂Cl) C, H, N.

Ethyl 2-Cyano-2-acetamido-5-(N-tosyl-N-3-acetamidopropyl)aminopentanoate (20). To a suspension of sodium hydride (0.557 g of a 55% oil suspension, 1.16×10^{-2} mol) in anhydrous DMF (5 mL) cooled to 0 °C was added under N2 a solution of ethyl acetamidocyanoacetate (1.972 g, 1.16×10^{-2} mol) in anhydrous DMF (15 mL). Stirring was continued at 0 °C for 1 h; then a solution of N-(3-chloropropyl)-N-tosyl-N'-acetyl-1,3-propanediamine (28) (4 g, 1.16×10^{-2} mol) in anhydrous DMF containing a catalytic amount of NaI was added over a period of 0.5 h. The reaction mixture was stirred overnight at room temperature and then heated at 100 °C for 1 h. The solvent was eliminated under reduced pressure and the residue dissolved in CHCl₃. The organic phase was washed with water and brine, dried over MgSO₄, and then concentrated under reduced pressure. Crystallization from AcOEt-EtOH yielded 2.4 g of 20 (45%): mp 94 °C. Anal. (C₂₂H₃₂O₆N₄S) C, H, N.

Ethyl 2-cyano-2-acetamido-5-phthalimidopentanoate (21) was obtained by the same procedure as above. N-(3-Iodopropyl)phthalimide was used in place of N-(3-chloropropyl)-N-tosyl-N-acetyl-1,3-propanediamine: overall yield from ethyl acetamidocyanoacetate, 62%; mp 206 °C. Anal. ($C_{18}H_{19}O_5N_3$) C, H, N.

1-(5-Tetrazolyl)-1,4-butanediamine Hydrochloride (10). A suspension of 21 (3.99 g, 1.15×10^{-2} mol), sodium azide (0.81 g, 1.25×10^{-2} mol), and ammonium chloride (0.69 g, 1.3×10^{-2} mol) in DMF (11 mL) was heated at 90 °C for 20 h. The solvent was eliminated under reduced pressure and the residue taken up in aqueous 0.5 M HCl (30 mL). The white precipitate which formed was collected by filtration, washed with water, and dried under vacuo over P₂O₅ to give 4.58 g of the tetrazolyl derivative 22 as an amorphous powder which was treated in concentrated HCl (40 mL) at reflux temperature for 72 h. The phthalic acid which precipitated on cooling was filtered off and the filtrate was concentrated under reduced pressure. The residue was purified by ion-exchange chromatography (Amberlite IR 120 H⁺ form, elution with 2 M NH₄OH). The fractions giving a positive test to ninhydrin were collected and concentrated to yield 1.17 g of the free base of 10. The monohydrochloride was prepared by adjusting the pH of an aqueous solution of the free diamine to 3.3 with 0.1 M HCl. Concentration of this solution and recrystallization (H₂O-EtOH-AcOEt) yielded analytically pure 10, mp 275 °C. Anal. $(C_5H_{13}N_6Cl)$ C, H, N.

DL-δ-N-3-Aminopropylornithine Dihydrochloride (8). A suspension of 20 (2 g, 0.42×10^{-2} mol) in concentrated HCl was heated at reflux temperature for 2 days. The residue obtained after evaporation of the solvent was passed through an Amberlite IR 120 H⁺ form column. Elution with 2 M NH₄OH and concentration of the fraction giving a positive test to ninhydrin afforded 0.79 g of a mixture of δ -N-3-aminopropylornithine and of its δ -N-tosylated analogue. This mixture was dissolved in liquid ammonia (150 mL) and sodium was added until a blue color persisted for at least 2 min. Ammonium chloride was added to discharge the color and ammonia was stripped off under a stream of N₂. The residue was passed through an Amberlite IR 120 H⁺ form resin column. The ninhydrin-positive fractions eluted with 2 M NH₄OH were collected and concentrated to give 0.47 g of crude δ -N-3-aminopropylornithine which was purified through recrystallization of its dipicrate salt. The dipicrate salt (1.19 g) was triturated with 1 M HCl (25 mL) for 0.5 h. The precipitate was collected by filtration and the filtrate was extracted with ether $(6 \times 20 \text{ mL})$ and then concentrated under reduced pressure. The residue was passed through an Amberlite IR 120 H⁺ form resin column. Elution with 2 M NH₄OH yield 230 mg of δ-N-3aminopropylornithine as an oil. The dihydrochloride was obtained by addition of HCl until a pH 4 was reached and was recrystallized from H_2O -EtOH (245 mg) (23%): mp 241 °C. Anal. (C_8H_{21} - $N_3O_2Cl_2$) C, H, N.

DL-δ-N-3-Aminopropyl-α-methylornithine Dihydrochloride (3). Treatment of 16c (1.55 g, 0.325×10^{-2} mol) in a manner similar to that described for 20 afforded 3 (390 mg, 45%), mp 245 °C. Anal. (C₉H₂₃N₃O₂Cl₂) C, H, N.

DL-δ-N-Methyl-α-methylornithine Hydrochloride Monohydrate (1). Treatment of 16a in a manner similar to that described for 20 afforded 1 (45%), mp 239 °C. Anal. (C_7H_{19} -N₂O₃Cl) C, H, N.

DL- α -Methyl- δ -N,N-dimethylornithine Dihydrochloride (2). A solution of 16b (2.1 g) in 6 M HCl was treated at 90 °C for 15 h. The solvent was concentrated under reduced pressure and the residue crystallized from ethanol to give 1.6 g of 2 (90%), mp 246 °C. Anal. $(C_8H_{20}O_2N_2Cl_2)$ C, H, N.

N-(3-Chloropropyl)-N-tosyl-N'-acetyl-1,3-propanediamine(28). A solution of N-(2-cyanoethyl)-N-tosyl-3-chloro-1propanamine (3.87 g, 0.129×10^{-2} mol) in acetic anhydride (120 mL) was hydrogenated over Platin Adams (400 mg) at atmospheric pressure. After the theoretical quantity of H2 has been absorbed, the catalyst was filtered off, and the filtrate was concentrated under reduced pressure. The residue was chromatographed on a silica gel column (200 g). Elution with AcOEt afforded 2.09 g of 28 (50%), oil. Anal. (C₁₅H₂₃N₂O₃SCl) C, H, N.

N-(2-Cyanoethyl)-N-tosyl-3-chloro-1-propanamine. To a solution of 3-(N-2'-cyanoethyl)amino-1-propanol²⁰ (6.83 g, 5.3 \times 10⁻² mol) in pyridine (10 mL) was added slowly 2 equiv of TsCl $(22.4 \text{ g}, 11.7 \times 10^{-2} \text{ mol})$. The stirring was continued for 24 h at room temperature. Recrystallization (Et₂O-pentane) of the residue obtained after evaporation of the solvent gave 12.2 g of N-(2'-cyanoethyl)-N-tosyl-3-chloro-1-propanamine (74%), mp 37 °C. Anal. (C₁₄H₁₉N₂SO₂Cl) C, H, N.

Benzyl α -isocyanopropionate (25) was prepared from Nformylalanine benzyl ester according to the method described by Walborsky³⁵ (65%): bp 94–96 °C (0.1 mmHg). Anal. $(C_{11}H_{10}O_2N)$ C, H, N.

Benzyl 2-Isocyano-2-methyl-5-(N,N-dimethylamino)pentanoate (16b). A suspension of NaH (0.02 mol) in THF (7.5 mL) was added dropwise under N2 to a well-stirred solution of 25 (3.74 g, 0.02 mol), N,N-dimethyl-3-chloro-1-propanamine (2.57 g, 0.02 mol), and NaI (catalytic amount) in a mixture of anhydrous Me₂SO (10 mL) and THF (10 mL). After the evolution of H₂ had stopped, the reaction mixture was treated at reflux temperature for 0.5 h, allowed to cool to room temperature, and then poured into ice water. Extraction with CH₂Cl₂ (4 × 60 mL) gave, after evaporation of the solvent, an oily residue (4.5 g) which was chromatographed on a silica gel column (250 g). Elution with AcOEt-EtOH (3:1) afforded 2.4 g of 16b (44%): oil; IR (film) 2140 (N=C), 1750 cm⁻¹ (CO₂CH₂Ph); NMR (CDCl₃) 1.66 (3 H, s, CCH₃), 2.18 [6 H, s, N(CH₃)₂], 5.25 (2 H, s, -OCH₂Ph), 7.4 (5 H, s, H aromatic); no analysis.

Benzyl 2-isocyano-2-methyl-5-(N-methyl-N-tosyl)aminopentanoate (16a) was obtained from N-methyl-N-tosyl-3-iodo-1-propanamine 18 in a manner similar to that described for 16b (74%), an oil. Anal. $(C_{22}H_{26}N_2O_4S)$ C, H, N.

Benzyl 2-isocyano-2-methyl-5-(N-tosyl-N-3-acetamidopropyl)aminopentanoate (16c) was obtained from 28 in a manner similar to that described for 16b (30%): an oil; no analysis.

(-)-5-(3-Aminopropyl)-5-methyl-2,4-imidazolinedione (30). A suspension of $(+)-\alpha$ -N-carbamoylbenzyloxycarbonyl- α methylornithine (150 mg) $[\alpha]^{20}$ +3.8° (c 0.45, MeOH), prepared by treating $(+)-\delta$ -N-benzyloxycarbonyl- α -methylornithine with an excess of KNCO in 1 M HCl (5 mL)] was heated at 90 °C for 4 h. The homogeneous reaction mixture was then concentrated in vacuo. The amorphous residue was dissolved in EtOH (5 mL) and the solution was stirred under an H₂ atmosphere for 3 h in the presence of 5% Pd/C (25 mg). Elimination of the catalyst by filtration and recrystallization from absolute EtOH (three times) yielded 22 mg of 30: mp 208-209 °C; $[\alpha]^{20}$ _D -14.5° (c 0.42, MeOH). Anal. $(C_7H_{13}N_3O_2)$ C, H, N.

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